Table 2. Checklist of essential features related to elemental analysis of biological systems

Specimen	Required information	
Plasma	Amount and type of anticoagulant and its chemical purity	
Serum	Body position during sampling, hemolysis status	
Milk	Days past partum, specific fraction (hindmilk, foremilk) if entire volume not collected	
Cells	Viability, (cell age), amount. Type and purity of stabilizer used, trapped plasma	
Soft tissues	Residual blood, decidual tissue while handling placenta, biopsy or autopsy	
Hard tissues		
bone	Biopsy or autopsy, renal function status sampling location	
hair and nail	Origin and washing procedures for hair and nail	
Diets	Proximate composition, caloric energy	
Feces	Occult blood	

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Ultra-Trace Elemental an	d Isotopic
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Trace element accumulation in the human fetus occurs primarily in the third trimester of pregnancy, and premature birth interrupts this process. A study of zinc in low birth weight infants indicates that the fetus accrues 310  $\mu$ g of Zn daily at the 30th week, increasing to 590 µg daily by the 36th week of gestation [1]. Similarly, the human fetus accumulates 80-90 µg of Cu/kg/day between 28 and 36 weeks, and by 40 weeks the fetus has accumulated almost 20 mg of Cu, one half of which is in the liver [2]. These and other essential trace elements play vital roles in the adult. Zinc is required for the synthesis of DNA, RNA, and protein, and as the zinc metallo-enzyme, regulates growth through DNA polymerase, RNA polymerase and thymidine kinase. References to these effects have been summarized [3]. Significant progress has been achieved recently using stable isotope tracers to assess the metabolic and nutritional roles of Ca [4], Zn [5], Se [6], and Mg [7] in adults and healthy infants. Techniques used for these studies include mass spectrometry, using electron impact ionization of metal chelates or thermal ionization of inorganic species. Trace elements at higher concentrations or in large samples have been determined with techniques such as atomic absorption (AA) or emission spectroscopy that do not address measurement needs for trace element and isotope tracer determinations in premature infants, healthy newborns, children, pregnant women, and other adults.

Little is known about the presence and function of trace and ultra-trace elements in pre-term infants whose birth weight is 500–2000 g. Only microliters of blood are available, and trace elements can occur in the blood at levels of parts per billion (ng/g)or lower. For a trace element level of 1 ppb, the total amount of the element available is 100 pg, or assuming vanadium, only  $10^{12}$  atoms. Clearly a new and comprehensive method is required to access elemental and isotopic information at these levels. Resonance ionization spectroscopy (RIS) is being used to solve these problems.

RIS utilizes a source of tunable laser radiation that is resonant with a specific atomic energy level. In the simplest two photon RIS scheme (fig. 1, scheme 1), a second photon of the same energy promotes the excited electron to the ionization continuum, forming an atomic ion. This simple two photon process can be used to ionize approximately fifty elements, but a series of RIS schemes has been proposed by Hurst et al., to ionize all elements except He and Ne (fig. 1) [8].

Resonance ionization advantages are threefold: 1) sensitivity, 2) selectivity, and 3) generality. Ionization sensitivity is achieved by saturating the photon absorption cross section of the energy level such that every atom crossing the active ionizing laser volume is ionized with unit probability. Elemental ionization selectivity is achieved through wavelength tunability. By selecting appropriate non-overlapping wavelengths, elements can be measured quantitatively in the presence of  $10^6 - 10^{12}$  of other elements. Resonance ionization is a general ionization process as described above.

# Experimental

The experimental design was structured toward the longer term research goal: to provide for the neonatologist a total multi-elemental and isotopic diagnostic analysis using only a few microliters of blood. Thus, the chemical separations must be minimal, simple, and capable of providing a group separation in a form amenable to RIS analysis. We selected electrodeposition as an initial separation process and were successful in depositing nanogram or picogram quantities of Cu, Mo, Se, and V singly or simultaneously onto a high purity gold substrate directly from aqueous solution. Cu and Mo were deposited individually onto gold from processed serum samples and their concentrations determined by isotope dilution. To separate copper, the serum samples were wet-ashed with a

nitric-perchloric acid mixture, evaporated to dryness, adjusted to the appropriate acidity, and the copper electrodeposited. To separate molybdenum, the sample was wet-ashed as above but the molybdenum was deposited from alkaline solution.

Following the multicomponent analysis theme, the laser and optical systems in Atom Sciences' Sputter-Initiated Resonance Ionization Spectrometer (SIRIS) (fig. 2) were modified to permit access to Cu, Zn, Mo, Se, and V within a single tunable laser dye range. Samples on gold foil were atomized by ion sputtering and the diluted isotope ratios were determined using a double-focusing mass spectrometer. The mass spectrometer was calibrated using gravimetrically prepared mixtures of isotopically natural copper and enriched <sup>65</sup>Cu.

## **Results and Discussion**

The results of the SIRIS calibration are illustrated in figure 3a. The data points cover an isotope ratio range of nearly 600, from natural copper ( $^{63}Cu/^{65}Cu=2.244$ ) to the separated  $^{65}Cu$  isotope ( $^{63}Cu/^{65}Cu=0.0038$ ). This range encompassed the anticipated  $^{63}Cu/^{65}Cu$  range for the isotope dilution samples, nominally 0.05 to 0.4. The deviation of each observed ratio from its gravimetric value was summed over all the ratios, yielding a relative standard deviation (RSD) of 4.7%. Isotope ratio precisions internal to an analysis were typically 1–2%, with attendant standard errors of the mean of 0.2%.

The concentration of copper was determined in separately processed duplicate samples of human serum and plotted in figure 3b versus analyses of the same (but larger) samples by flame AA. The RIS sample sizes were  $80-200 \ \mu\text{L}$ , and an overall accuracy of  $\pm 10\%$  was estimated, which included contributions from inhomogeneities, RIS methodology and the flame AA results using larger (2-3 mL) aliquots. Using isotope dilution with <sup>100</sup>Mo, Mo was determined in high purity reagents and serum. Concentrations of Mo in reagents, pg/g, were: H<sub>2</sub>O, 13; HNO<sub>3</sub>, 44; HC1O<sub>4</sub>, 30; and an Mo concentration of 19 ng/g was determined for a 250  $\mu$ L aliquot of Bovine Serum (RM 8419), compared to the suggested value of 16 ng/g [9].

Using SIRIS, sensitivities have been observed for Mo isotopes at the few pg level. Overall ionization and instrumental transmission efficiencies indicate potential absolute sensitivites of a few thousand atoms ( $\leq 10^{-9}$  monolayer). The RIS elemental and isotopic analysis technology is expected

to be generally applicable in medicine and nutrition for metabolic and diagnostic studies.

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Figure 1. A schematic illustration of five resonance ionization processes that are possible using commercially available lasers (taken from Ref. [8]). RIS of Mo is by a generalization of scheme 1, where  $\omega_1$  refers to a frequency doubled laser beam. Analogously, RIS of Cu is by a generalization of scheme 3, where  $\omega_1=\omega_2=$  frequency doubled beams, although ionization is by  $\omega_3=$  IR.



Figure 2. The sputter-initiated resonance ionization spectrometer (SIRIS) system used for the analytical determinations reported here. Condensed phase samples are atomized with an energetic argon primary ion beam ionized with a multi-color tunable dye laser system, and the ions are energy and mass analyzed with a double focusing mass spectrometer.



Figure 3. a. A calibration of the SIRIS system using <sup>63</sup>Cu/<sup>65</sup>Cu isotope ratio standards, prepared by mixing natural copper ( $^{63}$ Cu/ $^{65}$ Cu=2.244) and the  $^{65}$ Cu enriched isotope ( $^{63}$ Cu/ $^{65}$ Cu=0.0038). The relative standard deviation observed for the calibration was 4.7%; b. a plot of copper concentrations in bovine (RM 8419) and human serum determined with SIRIS, versus concentrations determined by flame AA using 2-3 mL samples. Serum sample sizes used for SIRIS were nominally 100  $\mu$ L and were separate duplicate determinations. The lowest concentration reported here was for bovine serum (RM 8419), and was 0.76  $\mu$ g/g compared to the suggested value of 0.73  $\mu$ g/g (Ref. [9]).

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# Use of High Resolution GC/MS for Obtaining Accuracy in Lipid Measurements

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## Introduction

Inaccuracy in clinical chemistry measurements is a serious problem in both service and research laboratories. It is only by having procedures of proven accuracy available that the clinical chemist can gain assurance that a working method is providing meaningful data. Analytical uncertainties occur in all areas of clinical chemistry and two examples of such problems in the area of lipid analysis are discussed here.

#### **Prostanoid Measurements**

One excellent example in which difficulties with inaccuracy can be seen is in the measurement of prostanoids. Although these analyses are not yet routine clinical chemistry measurements, these metabolites of arachidonic acid are of great importance in a vast number of research studies. Disturbances of prostanoid production are seen in a wide variety of pathological conditions including diabetes mellitus, hypersensitivity and inflammation, cancer, reproductive problems, hypertension and cardiovascular disease. To emphasize the analytical problems associated with measuring prostanoids, a survey of the literature reveals an incredible spread of reported reference (normal) ranges for various prostanoids in human plasma. For example, reported reference ranges for 6-keto-PGF<sub>1a</sub> are 0.4-5.0 pg/mL [1], 19-25 pg/mL [2], 160-228 pg/mL [3] and 1500–1700 pg/mL [4].

We have applied gas chromatography/mass spectrometry (GC/MS) to the measurement of prostanoids in plasma and in urine. Our procedure initially used an initial extraction, clean-up by high performance thin layer chromatography [1], derivatization, and detection by GC/MS at low or unit mass resolution using negative ion chemical ionization. Using this technique, assays of marginal performance were developed for the measurement of PGE<sub>2</sub>, 6-keto-PGF<sub>1a</sub>, PGF<sub>2a</sub>, and TxB<sub>2</sub> in plasma. However, when urine was analyzed, there were many interferences due to the complex matrix and reliable analytical data could not be produced.

The purpose of the present investigation was to reduce these potential interferences from biological specimens, particularly urine, by applying stable isotope dilution with quantitation by capillary column gas chromatography and high resolution mass spectrometry. The use of high resolution mass spectrometry is notable because it provides the selectivity of detection previously obtained by extensive chemical clean-up. This selectivity of detection can result in lower limits of detection despite a reduction in absolute sensitivity since one source of noise, chemical noise, is reduced more rapidly.

#### **Materials and Methods**

Deuterium-labeled internal standards  $(d_4-PGE_2$ and  $d_4$ -6-keto-PGF<sub>1a</sub>) were added to 10 mL of acidified urine (pH=2.5). Initial extraction of the prostanoids was by a C<sub>18</sub> SEP-PAK extraction, elution and subsequent ethyl acetate extraction from